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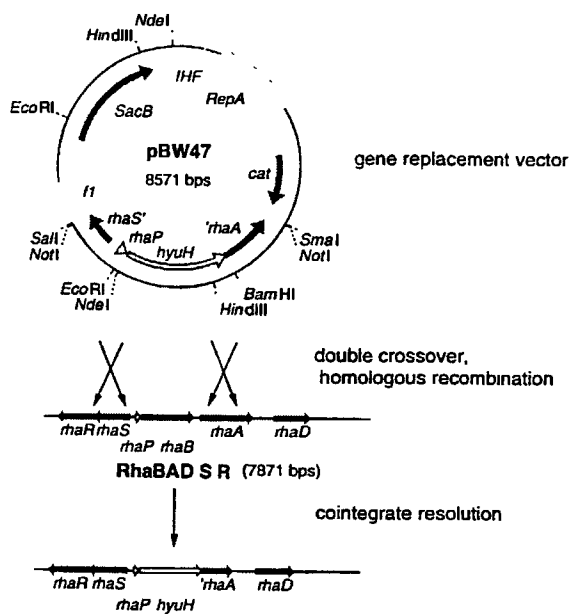
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(54) Title: WHOLE CELL CATALYST COMPRISING A HYDANTOINASE, A RACEMASE AND A CARBAMOYLASE

Chromosomal insertion of *hyuH*



(57) Abstract: A whole cell catalyst is described comprising a hydantoinase, a racemase and a carbamoylase. Thus this catalyst is able to degrade hydantoins directly into the amino acids. Additionally, a process for the production of this catalysts and for the production of amino acids is claimed.



— *With (an) indication(s) in relation to deposited biological material furnished under Rule 13bis separately from the description.*

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WHOLE CELL CATALYST COMPRISING A HYDANTOINASE, A RACEMASE AND A CARBAMOYLASE

The present invention is directed to a micro-organism, which is able to degrade hydantoins to enantiomerically enriched amino acids. Especially, this micro-organism is
5 equipped with cloned genes coding for the necessary enzymes.

Racemic 5-monosubstituted hydantoins can be chemically synthesized according to Bucherer-Berg method using aldehydes, ammonium bicarbonate and sodium cyanide as
10 reactants. They are important precursors for the enzymatic production of D-and L- amino acids. With the increasing demand for optically pure amino acids a lot of effort has been made towards the isolation of microorganisms capable for stereospecific hydrolysis of the hydantoins and
15 characterization of the enzymes (Syldatk and Pietzsch, "Hydrolysis and formation of hydantoins" (1995), VCH Verlag, Weinheim, pp. 403-434; Ogawa et al., J. Mol. Catal. B: Enzym 2 (1997), 163-176; Syldatk et al., Appl. Microbiol. Biotechnol. 51 (1999), 293-309). The asymmetric
20 bio-conversion to either L- or D- amino acids consists of three steps:

- (i) chemical and/or enzymatic racemization of 5-substituted hydantoins
- (ii) ring opening hydrolysis achieved by a hydantoinase
25 and
- (iii) hydrolysis of the N-carbamoyl amino acid produced by hydantoinase to the amino acid by carbamoylase.

Arthrobacter aurescens DSM 3747 is one of the few isolated microorganisms capable of converting 5-monosubstituted
30 hydantoins to L-amino acids. The disadvantage of using A. aurescens cells as biocatalyst is the low enzyme activity. Especially the L-N-carbamoylase is the bottleneck for most

substrates leading to an increase of the intermediate L-N-carbamoyl amino acid in the cell, which is not further converted to the corresponding amino acid. By combining the purified enzymes bottlenecks could be avoided but due to
5 the low amounts of enzymes in the cells and loss of activity during the many necessary purification steps this process is not cost-effective.

All three genes encoding for the racemase hyuA (seq. 11), the L-specific hydantoinase hyuH (seq. 9) and the
10 stereoselective L-N-carbamoylase (seq. 7) have been cloned in *E. coli* separately and expressed to high levels (about 10 % of the total cell protein) (DE 19913741; J. Biotechnol., to be published). For in vitro catalysis the enzymes from the three recombinant strains can be produced
15 and purified more cost-effective than from the *Arthrobacter aurescens* strain. Regarding the different enzyme activities towards the various substrates the enzymes can be combined in enzyme reactors at ratios optimized for each reaction.

It is an object of this invention to provide a further
20 possibility of how a racemase, a hydantoinase and a D- or L-specific carbamoylase can act together in a process for the production of enantiomerically enriched amino acids from 5-monosubstituted hydantoins. Especially, this possibility should be suitable to be implemented in
25 processes on technical scale, that is to say it has to be most cost-effective.

This is done by using a whole cell catalyst according to claim 1. Further preferred catalysts are subjects to claims depending from claim 1. Claims 6 to 9 are directed to a
30 process for the production of the whole cell catalyst of the invention. Claims 10 and 11 protect a process for the production of enantiomerically enriched amino acids using the catalyst according to the invention.

Using whole cell catalysts comprising cloned genes encoding for a hydantoinase, for a hydantoin racemase and a D- or L-specific carbamoylase for the conversion of 5-monosubstituted hydantoins to L- or D-amino acids results in a fast and complete conversion of racemic mixtures of hydantoins to the corresponding L- or D-amino acids on industrial scale. This significantly reduces the production costs due to a reduction of fermentation and purification costs because all enzymes are produced in one strain.

Advantageously, a bacteria is used as cell, because of high reproduction rates and easy growing conditions to be applied. There are several bacteria known to the skilled worker which can be utilized in this respect. Preferably a *Escheria coli* can be used as cell and expression system in this regard (Yanisch-Perron et al. Gene (1985), 33, 103-109).

It is another positive embodiment of this invention that in principle all genes encoding for the hydantoinase, racemase and carbamoylase known to the artisan can be taken to be expressed in the whole cell catalyst. Preferably all genes can be taken from DSM 3747 (seq. 7, 9, 11).

The enzymes to be incorporated in the genetic code of the whole cell catalyst naturally possess different turnover rates. It is a drawback if the rates of co-working enzymes are not in line and intermediates accumulate during the production inside the cell. The overexpression of the hydantoinase gene in *E. coli* leads to the formation of inclusion bodies (Wiese et al., in preparation), which is unfavourable for a well balanced coexpression of all the three enzymes. Therefore, various attempts to "fine tune" the expression of these genes have been made. This can be done advantageously by overexpressing the hydantoinase genes in question according to their turnover rates. According to the DSM 3747-System the hydantoinase gene is overexpresses from plasmids with reduced copy numbers.

A further embodiment of the instant invention is directed to a process for the production of the whole cell catalyst according to the invention. In principle all plasmids known to the skilled worker can serve to carry the gene into the expression system. Preferably, plasmids derived from pSC101, pACYC184 or pBR322 are used to produce the catalyst. Most preferably plasmids pBW31 and pBW32, pBW34 and pBW35, pBW34 and pBW53, pBW32 or pBW34 are used in this respect. For the skilled worker plasmids and methods to produce plasmids can be deduced from Studier et al., Methods Enzymol. 1990, 185, 61-69 or brochures of Novagen, Promega, New England Biolabs, Clontech or Gibco BRL. More applicable plasmids, vectors can be found in: DNA cloning: a practical approach. Volume I-III, edited by D. M. Glover, IRL Press Ltd., Oxford, Washington DC, 1985, 1987; Denhardt, D. T. and Colasanti, J.: A survey of vectors for regulating expression of cloned DNA in E. coli. In: Rodriguez, R.L. and Denhardt, D. T (eds), Vectors, Butterworth, Stoneham, MA, 1987, pp179-204; Gene expression technology. In: Goeddel, D. V. (eds), Methods in Enzymology, Volume 185, Academic Press, Inc., San Diego, 1990; Sambrook, J., Fritsch, E.F. and Maniatis, T. 1989. Molecular cloning: a laboratory manual, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. They are incorporated by reference herewith.

Over-expression can be accomplished by means known to the skilled artisan, e.g. using constitutive or inducible expression systems as reviewed by Makrides (Makrides, 1996, Microbiol. Rev. 60, no. 3, 512-538)

Preferably, for expression of the enzymes a rhamnose inducible E. coli promoter cassette is used.

In addition, primers useful for the amplification of the gene of the invention in a PCR are protected similarly. Primers which are feasible are for example, primers S988 (seq. 6), S2480 (seq. 1), S2248 (seq. 2), S2249 (seq. 3),

S2517 (seq. 4) or S2518 (seq. 5). Furthermore, all other primers which could serve to carry out this invention and which are known to the artisan are deemed to be useful in this sense. The finding of a suitable primer is done by
5 comparison of known DNA-sequences or translation of amino acid sequences into the codon of the organism in question (e.g. for Streptomyces: Wright et al., Gene 1992, 113, 55-65). Similarities in amino acid sequences of proteins of so called superfamilies are useful in this regard, too
10 (Firestine et al., Chemistry & Biology 1996, 3, 779-783). Additional information can be found in Oligonucleotide synthesis: a practical approach, edited by M.J. Gait, IRL Press Ltd, Oxford Washington DC, 1984; PCR Protocols: A guide to methods and applications, edited by M.A. Innis,
15 D.H. Gelfound, J.J. Sninsky and T.J. White. Academic Press, Inc., San Diego, 1990. Those strategies are incorporated by reference herewith.

Another aspect of the invention is a process for the production of enantiomerically enriched amino acids, which
20 utilizes a whole cell catalyst according to the invention. Furthermore, a process is preferred that is performed in an enzyme-membrane-reactor (DE 19910691.6).

To adopt the turnover rate of all enzymes expressed in the whole cell catalyst to each other there are different
25 methods to achieve this.

a) The genes are expressed with different promoters. The gene with the lowest activity is combined with the strongest promoter and vice versa. A disadvantage would be that for each gene a different inductor is necessary to
30 induce the expression of all genes.

b) The genes are expressed with one promoter on a polycistronic messenger. The ratios of synthesis of the enzymes is influenced by changing or by mutation of the translation initiation region of each gene (the ribosomal

binding site) which determines the efficiency of protein synthesis. This principle is realized in operons of microorganisms. The disadvantage, is that the efficiency of a translation initiation region can not be predicted which means that for each gene many changes in the translation initiation region have to be made and tested (Grifantini et al., 1998, Microbiology, 144, 947-954).

c) The enzyme activity of each enzyme can be changed by mutation using error prone PCR (Fromant et al., 1995, Anal. Biochem. 224, 347-353) and DNA shuffling (Stemmer, 1994, Nature 370, 389-391). Again, this is very time consuming and costly.

d) Instead of mutagenizing genes to optimize their function in a reaction cascade, genes from different origins which encode enzymes with appropriate properties could be combined to an operon. This needs a large database describing such enzymes.

e) All genes are expressed from the same promoter but from replicons with different copy numbers. This can be the chromosome (single copy) or plasmids with low, moderate and high copy numbers. By constructing various compatible plasmids with different copy numbers and antibiotic markers carrying each the same cassette with the promoter and a polylinker sequence, genes of interest can be integrated into the plasmids in one step and the plasmids combined within one strain. This method allows a fast construction and testing of many combinations and with just one inducer in one fermentation all genes are expressed at different levels according to the plasmid copy number.

The following paragraphs show the transformation of Hydantoins to enantiomerically enriched amino acids.

Expression of the *hyuA* gene (seq. 11) is necessary for complete substrate conversion. Figure 1 shows the time

course of conversions with E.coli BW3110 containing pAW229 and pBW31. pAW 229 contains the carbamoylase gene on a pACYC plasmid, pBW31 is a pBR derivative and carries the hydantoinase gene. After consumption of 50% of the substrate, the reaction almost stops completely, since spontaneous racemisation of IMH is very slow (Syldatk et. al., "Biocatalytic production of amino acids and derivatives" (1992), Hanser publishers, New York, pp. 75-176). As can be seen from figure 2, bringing the racemase (seq. 11) into the system by using pBW31 and pBW32, the pACYC plasmid with the carbamoylase and the racemase gene, enables complete conversion of the substrate. After 4.5 hours induction at 30°C 200 µl permeabilized cells were prepared as described above and were incubated with 800 µl of 2 mM D,L-IMH.

The E. coli strain BW3110H with the chromosomally integrated hydantoinase gene was transformed with pBW32 (Figure 3), the pACYC plasmid containing the carbamoylase and the racemase gene, or with pBW34 (Figure 4), the pBR plasmid containing the carbamoylase and the racemase gene. Cells were induced at 25°C for 8.5 hours (pBW32), or for 11.5 hours (pBW34). Cell harvesting and permeabilization took place as described above.

Figure 5 shows E. coli BW3110 cells with pBW31 (the pBR plasmid which carries the hydantoinase gene) and pBW32 (the pACYC plasmid with the carbamoylase and racemase genes). Cells were induced 10 hours at 30°C.

The combination of pBW31 and 32 enables fast and complete conversion from D-L-IMH to tryptophane. The intermediate is formed up to a concentration of 0.4 mM.

E. coli BW3110 transformed with the plasmids pBW34 (the pBR plasmid with the carbamoylase and the racemase genes) and pBW35 (the pACYC plasmid with the hydantoinase gene) was taken for this conversion. Cells were induced for 10 hours at 30°C (Fig. 6). The combination of pBW34 and 35 shows an

accumulation of the intermediate up to over 1mM. The product formation takes place at a lower rate than seen in figure 5.

Plasmid pBW34 (the pBR plasmid with the carbamoylase and
5 the racemase genes) was combined with pBW53 (the pSC101
plasmid with the hydantoinase gene). Induction took place
for 10 hours at 30°C (Fig 7). In this case a fast
conversion of IMH to tryptophane takes place. Formation of
10 the intermediate is strongly reduced and product formation
is faster than seen in figure 5, so that the combination of
pBW34 and pBW53 is most favourable for the process.

The present invention shows a new and superior way to
combine a hydantoinase, a hydantoin racemase and a
carbamoylase in a whole cell catalysator. It is this
15 possibilty that renders instant invention to a proper
method for the production of enantiomerically enriched
amino acids from hydantoins due to reduction of catalyst
production costs.

Enantiomerically enriched means that one antipode of a
20 chiral compound is the major component in a mixture of both
antipodes.

Amino acid denotes within the framework of this invention
all compounds comprising a primary amine function connected
to a carboxylic acid group via one intermediate C-atom (α -
25 C-atom). This α -C-atom bears only one further residue.
Nevertheless all natural and unnatural amino acids are
deemed to be encompassed. Preferred unnatural amino acids
are those mentioned in DE 19903268.8.

Genes encoding for a peptide sequence are to be understood
30 as all genes possible with regard to the degeneration of
the genetic code.

The microorganism DSM 3747 is disposed at Deutsche
Sammlung für Mikroorganismen und Zellkulturen.

Examples:

Bacterial strains, plasmids and growth conditions:

E. coli JM109 (Yanisch-Perron et al. Gene (1985), 33, 103-109) was used for cloning procedures involving the hyuC (seq. 7), hyuH (seq. 9) and hyuA (seq. 11) genes from Arthrobacter aurescens DSM 3747 (Groß et al., Biotech. Tech. (1987), 2, 85-90). E. coli BW3110 (Wilms et al, in preparation), a derivative of E. coli W3110 (Hill and Harnish, 1981 Proc. Natl. Acad. Sci USA 78, 7069-7072) was used for coexpression for the genes mentioned above. E. coli strains were either grown in LB liquid medium or on LB-agar plates (Luria et al., 1960, Virology 12, 348-390), both supplemented with 100 µg/ml ampicillin and / or 25µg/ml chloramphenicol to select plasmid carrying strains. The cultures were grown at 37°C, for heterologous gene expression growth temperature was reduced to 30°C or 25°C.

General protocols:

All of the recombinant DNA techniques were standard methods (Sambrook et al., Molecular Cloning: A laboratory manual (1989), Cold Spring Harbour Laboratory Press, New York). PCR reactions were performed either with Pwo Polymerase or the ExpandTM Long Template PCR System by following the recommendations of Roche Diagnostics.

Coexpression of hyuA, hyuC, and hyuH in E. coli:

For coexpression of the racemase gene hyuA, the carbamoylase gene hyuC, and the hydantoinase gene hyuH in E. coli, several constructions with different features were made. To obtain comparable expression levels of the genes, variations in the copy number of plasmids were used. High copy plasmids like pBR plasmids (Bolivar et al., 1977, Gene 22, 277-288) have a copy number of 40-50. PACYC184 plasmids (Chang and Cohen, 1978, J. Bacteriol., 1141-1156) have a copy number of 10-15. PSC101 plasmids (Cohen et al., 1973, Proc. Natl. Acad. Sci. USA, 70, 3240-3244) have a copy

number of 5-10. A copy number of 1 is achieved by inserting the gene into the E. coli chromosome.

The plasmid features are summarized in table1:

plasmid name	ori	copy number	resistance	hyu - genes
pAW229	pACYC	10-15	cam	hyuC
pBW31	pBR	40-50	amp	hyuH
pBW32	pACYC	10-15	cam	hyuC + hyuA
pBW34	pBR	40-50	amp	hyuC + hyuA
pBW35	pACYC	10-15	cam	hyuH
pBW53	pSC101	5-10	cam	hyuH

5

Abbreviations: hyu: hydantoin utilizing

hyuA: racemase gene (seq. 11)

hyuC: carbamoylase gene (seq. 7)

hyuH: hydantoinase gene (seq. 9)

10

amp: ampicillin resistance (β -lactamase gene)

cam: chloramphenicol resistance
(chloramphenicol acetyl transferase gene)

15

The hydantoinase gene *hyuH* was also expressed using the strain BW3110H, which carries a chromosomal insertion of the *hyuH* gene.

5 All constructs enable transcriptional regulation of gene expression by the *rhaBAD* promoter.

For coexpression of the carbamoylase gene *hyuC* and the hydantoinase gene *hyuH* pAW229 and pBW31 are transformed into *E. coli* BW3110.

10 For coexpression of the racemase gene *hyuA*, the carbamoylase gene *hyuC* and the hydantoinase gene *hyuH*, pBW31 and pBW32, pBW34 and pBW35, or pBW34 and pBW53 are suitable combinations in *E. coli* BW3110.

To achieve coexpression of all three Enzymes in *E. coli* BW3110H, pBW32 or pBW34 can be used.

15 Construction of the plasmids:

pAW229 was obtained by cleaving pAW178 (Wilms et al., J. Biotechnol. (1999), 68, 101-113) with the restriction enzymes *NdeI* and *BamHI* and ligating the 1241bp fragment containing the *hyuC* gene into pJOE2962 (Altenbuchner, 20 unpublished), which was cut with the same restriction enzymes.

pBW31 was constructed by cleaving pAW92 (Wiese et al., in preparation) with the restriction enzymes *EcoRI* and *BamHI* and ligating the 1436bp fragment containing the *hyuH* gene 25 into pBW22, which was cut with the same restriction enzymes.

pBW32 was obtained by PCR amplification of the *hyuA* gene using the primers S988 (5'-AGGCTGAAAATCTTCTCT-3') (seq. 6) and S2480 (5'-AAAAAAGCTTTTAAAGAAGGAGATATACATA-3') (seq. 1) 30 and pAW210 (Wiese et al., in preparation) as template.

Included in primer S2480 is a shine dalgarno sequence for translation initiation. The fragment was inserted into the *HindIII* site of pAW229.

pBW34 was created by inserting the *hyuA* PCR fragment 35 described above into the *HindIII* site of pBW24. pBW24 was

obtained by cleaving pAW178 (Wilms et al, J. Biotechnol. (1999), 68, 101-113) with NdeI and HindIII and ligating the 1261bp long fragment containing the hyuC gene into pBW22, which was cut with the same restriction enzymes. pBW22 was
5 constructed by PCR amplifying of the "cer"-region from the colE1 plasmid using the primers S2248 (5'-AAA GCA TGC ATG GCC CTT CGC TGG GAT-3') (seq. 2) and S2249 (5'-AAA GCA TGC ATG GCT ACG AGG GCA-3') (seq. 3). The 268bp fragment was cut with the restriction enzyme SphI and inserted in the
10 vector pJOE2775 (Krebsfänger et al., 1998, Enzyme Microb. Technol. 22, 219-224) which was cut with the same restriction enzyme.

pBW35 was constructed by cleaving pBW31 with the restriction enzymes NdeI and BamHI. The 1379bp fragment
15 containing hyuH was inserted into pAW229, which was cut with the same restriction enzymes.

pBW53 was obtained by cleaving pBW31 with the restriction enzymes SphI and BamHI. The 1534bp fragment containing the hyuH gene and the rhamnose promoter was inserted into pSB27
20 (Baumann, Dissertation, Universität Stuttgart, 1996), which was cut with the same restriction enzymes.

Construction of the chromosomal integrate of hyuH into the rhamnose operon:

A 3.5kb fragment from the E. coli rhamnose operon was
25 amplified using the primers S2517 (5'-AAACAAGATCTCGCGACTGG-3') (seq. 4) and S2518 (5'-AAAAAGATCTTTATCAGGCCTACAACTGTTG-3') (seq. 5) and E. coli chromosomal DNA as template. The fragment was cut with the restriction enzyme BglII and inserted into the vector pIC20H (Marsh et al., 1984, Gene
30 32, 481-485), which was cut with the restriction enzymes BamHI and BglII, to get pBW39. PBW31 was cut with the restriction enzymes EcoRI and BamHI. The 1436bp fragment containing the hyuH gene was inserted into the vector pBW39, which was also cut with the same restriction
35 enzymes, to get pBW40. A 2.9kb fragment was amplified using the primers S2517 and S2518 and pBW40 as a template. This

fragment was cut with BglII and inserted into the vector pJOE2114 (Altenbuchner, unpublished) which was also cut with BglII to get pBW45. pBW45 was cut with BglII and SphI. The resulting 2.9kb *rhaS-rhaP-hyuH-rhaA* fragment was

5 inserted into the gene replacement vector pK03 (Link et al, 1997, J. Bacteriol., 179, 20, 6228-6237), which was cut with BamHI. The gene replacement was carried out according to the authors' instructions. Positive insertion events were screened using MacConckey Rhamnose plates.

10 Preparation of cells and activity measurements:
For induction of the *rhaBAD* promoter strains with two plasmids were grown in LB_{amp+cam}, strains with one plasmid in LB_{amp} or LB_{cam} respectively to OD₆₀₀ = 0.3-0.5. Then L-rhamnose was added to a final concentration of 0.1 g l⁻¹

15 and the cultivation was continued to a final OD of approximately 5. If not indicated separately, for small scale enzyme measurements cells corresponding to OD₆₀₀ of 20 were harvested, washed in 1 ml 0.2 M Tris pH 7.0 and resuspended in 1 ml 0.2 M Tris pH 7.0, 1 mM MnCl₂. 10 µl

20 toluene was added for permeabilizing the cell membranes. After 30min of incubation at 37°C 200 µl of this cell suspension were added to 800 µl of 2 mM D,L-Indolylmethylhydantoin (IMH) in 0.1 M Tris pH 8.5, mixed and shaken at 37°C. This cell amount corresponds to

25 approximately 5-6 mg cell wet weight. Samples were taken regularly. The reaction was stopped by adding 14% trichloroacetic acid. The time course of product and educt concentrations was determined using HPLC analysis. The HPLC-system was equipped with a RP-18 column as described

30 previously for the determination of hydantoin derivatives and N-carbamoyl amino acids (May et al., 1998, J. Biotechnol., 26, 61 (1): 1-13). UV-absorption was measured at 280 nm and the mobile phase (0.3% (v/v) phosphoric acid (80%) and methanol (20%; v/v)) was pumped with a flow rate

35 of 1.0 ml min⁻¹.

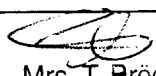
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0-3	Applicant's or agent's file reference	990160 AM
1	The indications made below relate to the deposited microorganism(s) or other biological material referred to in the description on:	
1-1	page	8
1-2	line	32
1-3	Identification of Deposit	
1-3-1	Name of depositary institution	DSMZ-Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH
1-3-2	Address of depositary institution	Mascheroder Weg 1b, D-38124 Braunschweig, Germany
1-3-3	Date of deposit	
1-3-4	Accession Number	DSMZ 3747
1-4	Additional Indications	NONE
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Claims:

1. Whole cell catalyst for the conversion of 5-monosubstituted hydantoins to L- or D-amino acids comprising cloned genes encoding for a hydantoinase,
5 for a hydantoin racemase and a D- or L-specific carbamoylase.
2. Catalyst according to claim 1, characterized in that a bacteria is used as cell.
- 10 3. Catalyst according to claim 1 and 2, characterized in that Escherichia coli is used as cell.
4. Catalyst according to one or more of preceding claims, characterized in that
15 the genes encoding for the hydantoinase, racemase and carbamoylase is taken from DSM 3747.
5. Catalyst according to one or more of preceding claims, characterized in that
20 the genes are overexpressed in the cell according to their turnover rates.
6. Process for the production of a whole cell catalyst, characterized in that plasmids derived from pSC101, pACYC184 or pBR322 are used.
- 25 7. Process according to claim 6, characterized in that plasmids pBW31 and pBW32, pBW34 and pBW35, pBW34 and pBW53, pBW32 or pBW34 are used.
8. Process according to claim 6, characterized in that an
30 E. coli strain with a chromosomally insertion of the hydantoinase gene is used.

9. Process according to claim 6,
characterized in that
a rhamnose inducible E. coli promoter cassette is
used.
- 5 10. Process according to claim 6,
characterized in that
primers S2480, S2248, S2249, S2517 or S2518 are used.
11. Process for the production of enantiomerically
enriched amino acids,
10 characterized in that
a whole cell catalyst according to claim 1 is used.
12. Process according to claim 11,
characterized in that
the process is performed in an enzyme-membrane-
15 reactor.
13. Process for the production of whole cell catalysts
characterized in that
all genes are expressed from the same promotor but
from plasmids with replicons with different copy
20 numbers.
14. Plasmids pBW31, pBW32, pBW34, pBW35, pBW53, AW229.
15. Primers S2480, S2248, S2249, S2517, S2518.
16. Microorganisms comprising plasmids according to claim
14.

AMENDED CLAIMS

[received by the International Bureau on 6 February 2001 (06.02.01);
original claims 1-16 replaced by new claims 1-15 (2 pages)]

1. Whole cell catalyst for the conversion of 5-monosubstituted hydantoins to L- or D-amino acids comprising cloned genes encoding for a hydantoinase, for a hydantoin racemase and a D- or L-specific carbamoylase, wherein the genes are overexpressed in the cell according to their turnover rates.
2. Catalyst according to claim 1, characterized in that a bacteria is used as cell.
3. Catalyst according to claim 1 and 2, characterized in that Escherichia coli is used as cell.
4. Catalyst according to one or more of preceding claims, characterized in that the genes encoding for the hydantoinase, racemase and carbamoylase is taken from DSM 3747.
5. Process for the production of a whole cell catalyst according to claim 1, characterized in that plasmids derived from pSC101, pACYC184 or pBR322 are used.
6. Process according to claim 5, characterized in that plasmids pBW31 and pBW32, pBW34 and pBW35, pBW34 and pBW53, pBW32 or pBW34 are used.
7. Process according to claim 5, characterized in that an E. coli strain with a chromosomally insertion of the hydantoinase gene is used.
8. Process according to claim 5, characterized in that a rhamnose inducible E. coli promoter cassette is used.

9. Process according to claim 5,
characterized in that
primers S2480, S2248, S2249, S2517 or S2518 are used.
10. Process for the production of enantiomerically enriched
amino acids,
characterized in that
a whole cell catalyst according to claim 1 is used.
11. Process according to claim 10,
characterized in that
the process is performed in an enzyme-membrane-reactor.
12. Process for the production of whole cell catalysts
according to claim 1
characterized in that
all genes are expressed from the same promotor but from
plasmids with replicons with different copy numbers.
13. Plasmids pBW31, pBW32, pBW34, pBW35, pBW53, AW229.
14. Primers S2480, S2248, S2249, S2517, S2518.
15. Microorganisms comprising plasmids according to claim 13.

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Fig 1:

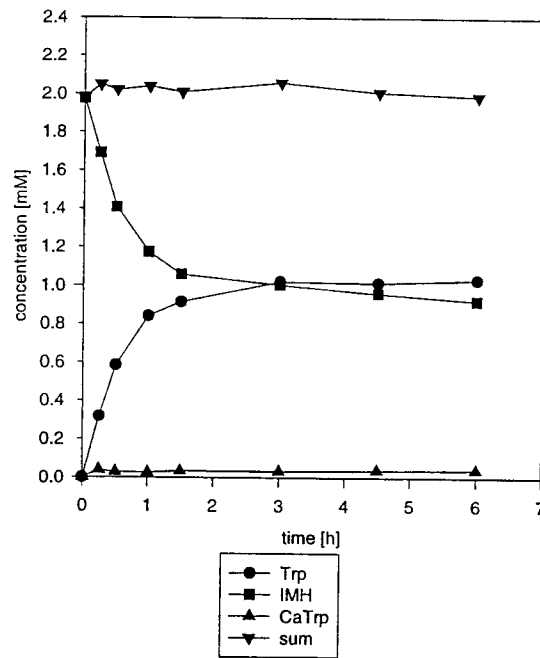
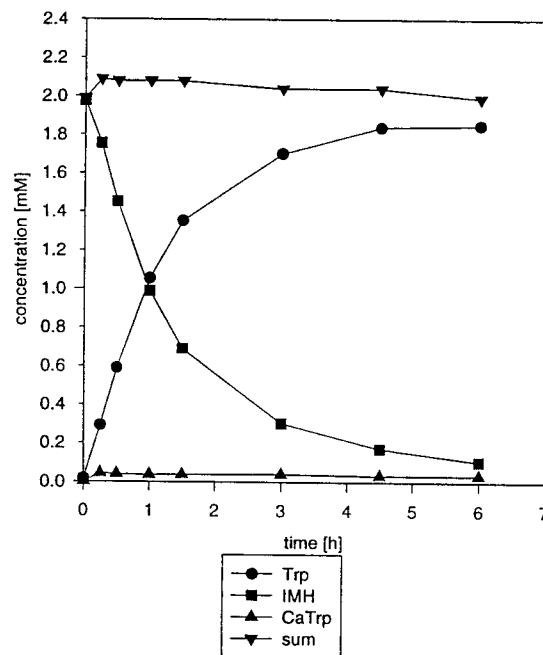


Fig. 2:



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Fig. 3:

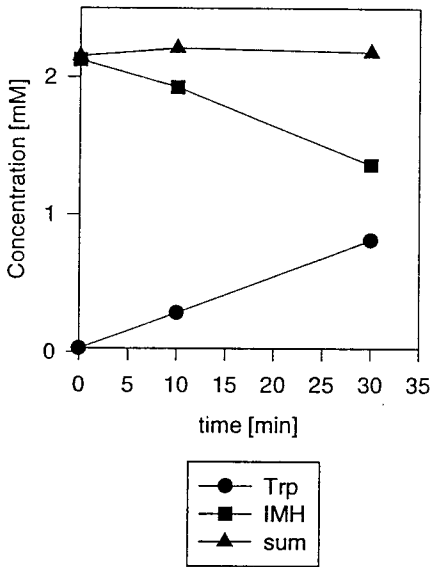
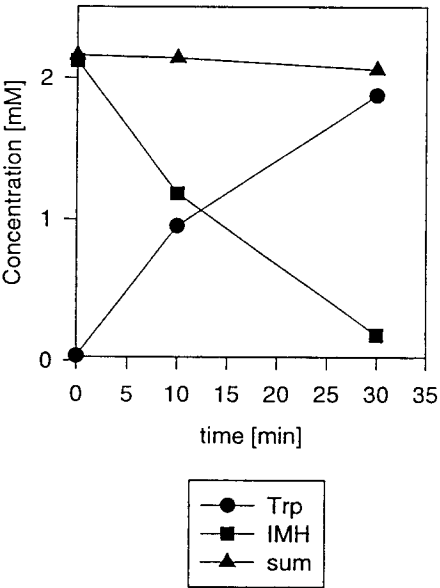


Fig. 4:



3110

Fig. 5:

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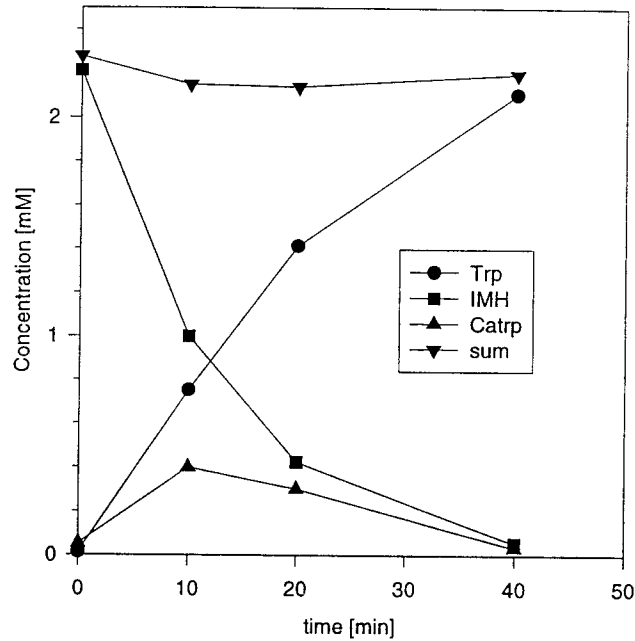
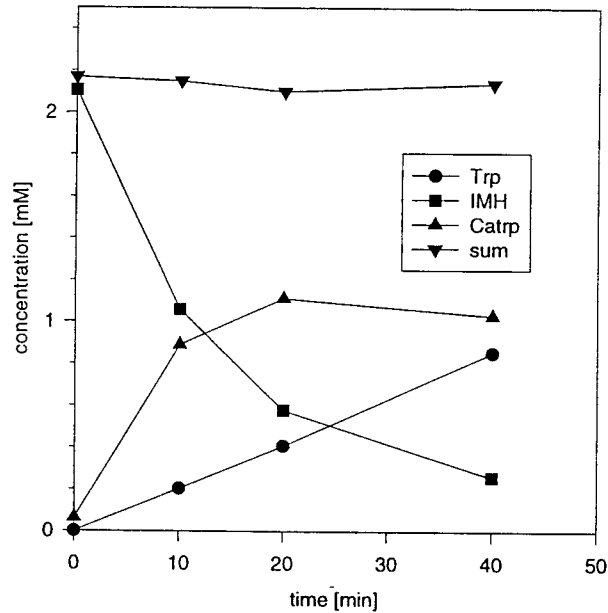


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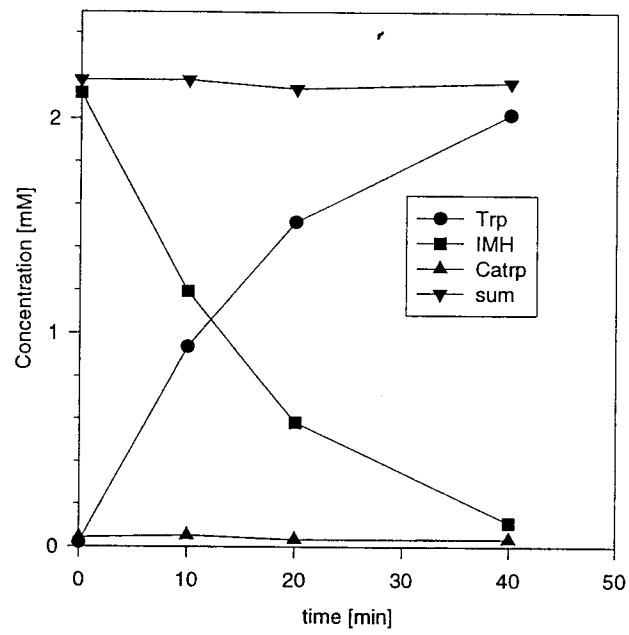
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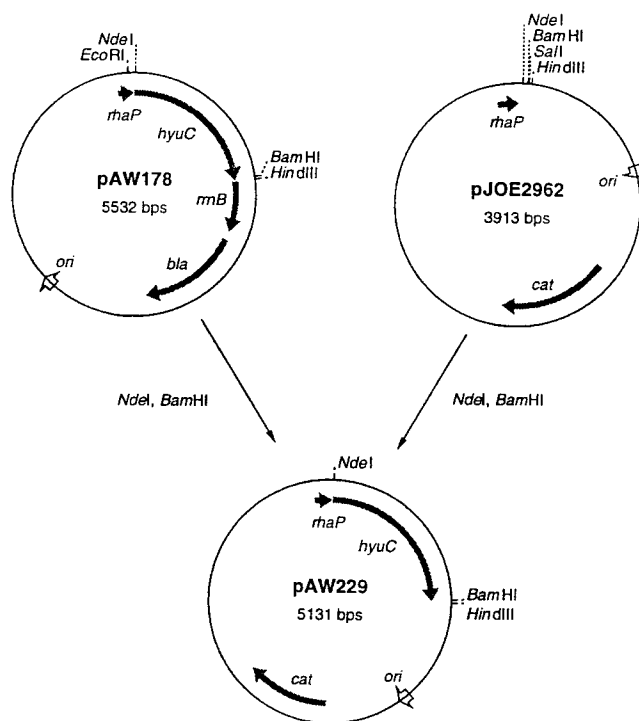
Fig. 7:



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Fig. 8:

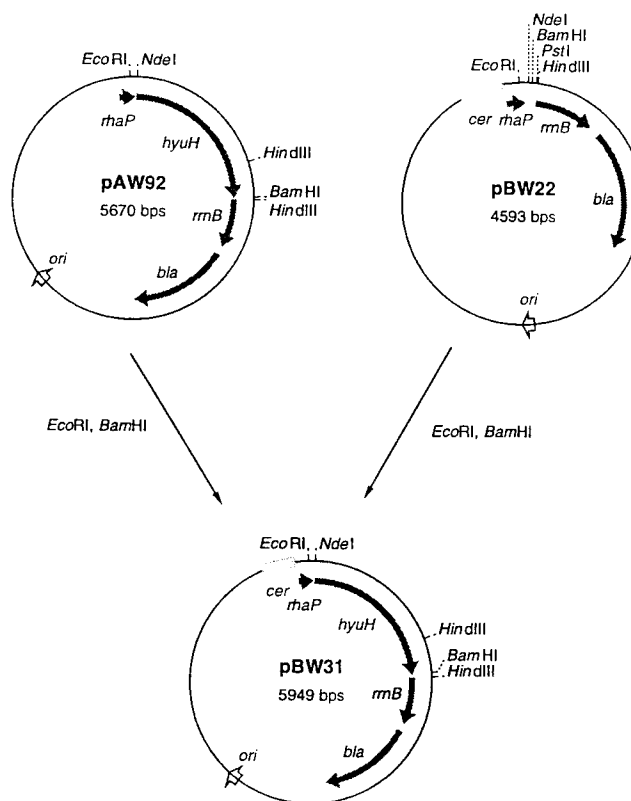
Construction of pAW229



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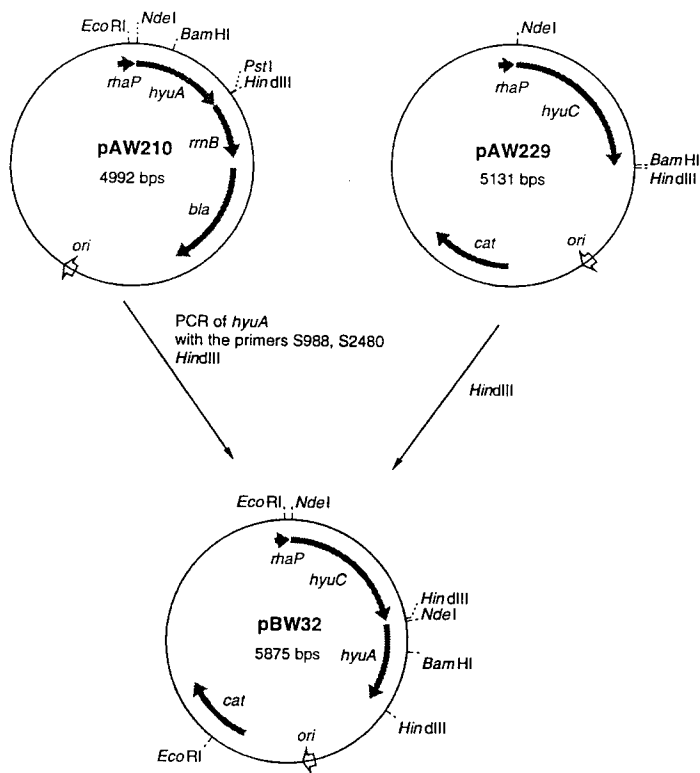
Construction of pBW31



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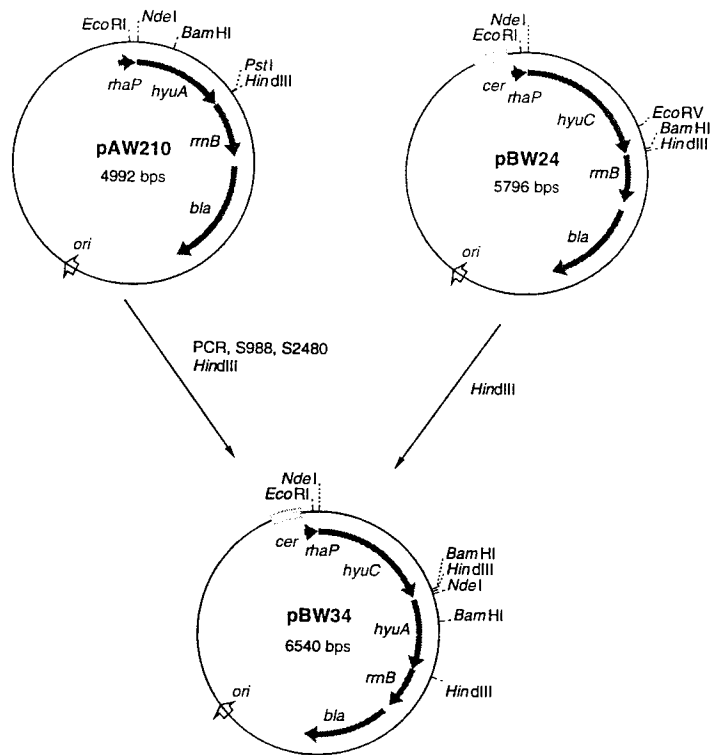
Construction of pBW32



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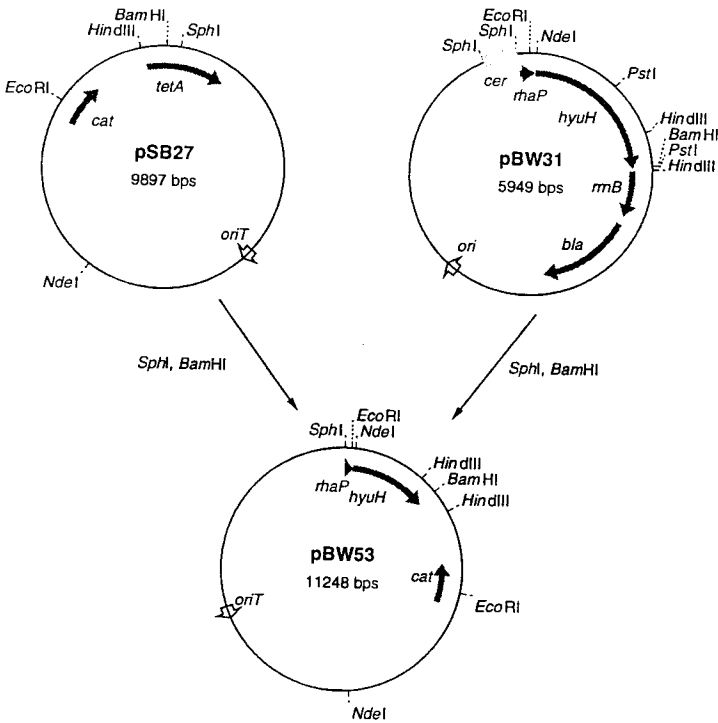
Construction of pBW34



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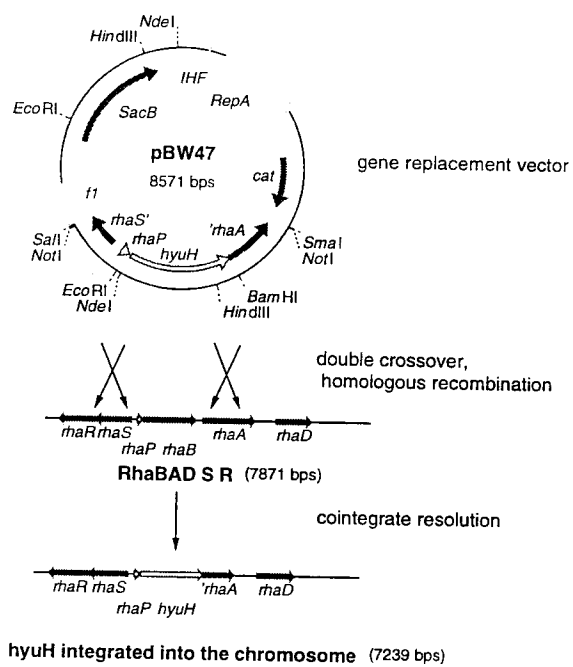
Fig. 12:

Construction of pBW53



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Fig. 13:

Chromosomal insertion of *hyuH*

1

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10	tcc att gtc agc atc ctc ccg agg gtc agg aaa cat ctg cac gaa ctg Ser Ile Val Ser Ile Leu Pro Arg Val Arg Lys His Leu His Glu Leu 115 120 125			384
	gta cgg caa gcg ggg gcg acg aat cgc ctc gcc tcc atc aag ctc cca Val Arg Gln Ala Gly Ala Thr Asn Arg Leu Ala Ser Ile Lys Leu Pro 130 135 140			432
15	aat ctg ggg gtg atg gcc ttc cat gag gac gaa cat gcc gca ctg gag Asn Leu Gly Val Met Ala Phe His Glu Asp Glu His Ala Ala Leu Glu 145 150 155 160			480
20	acg ctc aaa caa gcc gcc aag gag gcg gtc cag gag gac ggc gcc gag Thr Leu Lys Gln Ala Ala Lys Glu Ala Val Gln Glu Asp Gly Ala Glu 165 170 175			528
25	tcg ata gtg ctc gga tgc gcc ggc atg gtg ggg ttt gcg cgt caa ctg Ser Ile Val Leu Gly Cys Ala Gly Met Val Gly Phe Ala Arg Gln Leu 180 185 190			576
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	cgc gtg gcc gag agt ttg gtc gct ctg ggc tac cag acc agc aaa gcg Arg Val Ala Glu Ser Leu Val Ala Leu Gly Tyr Gln Thr Ser Lys Ala 210 215 220			672
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55	Asp Glu Ala Leu Ala Thr Phe His Leu Ile Glu Glu Val Glu Arg Ala 50 55 60			
	Glu Arg Glu Asn Pro Pro Asp Ala Tyr Val Ile Ala Cys Phe Gly Asp 65 70 75 80			

||

Pro Gly Leu Asp Ala Val Lys Glu Leu Thr Asp Arg Pro Val Val Gly
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5 Val Ala Glu Ala Ala Ile His Met Ser Ser, Phe Val Ala Ala Thr Phe
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Ser Ile Val Ser Ile Leu Pro Arg Val Arg Lys His Leu His Glu Leu
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10 Val Arg Gln Ala Gly Ala Thr Asn Arg Leu Ala Ser Ile Lys Leu Pro
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Asn Leu Gly Val Met Ala Phe His Glu Asp Glu His Ala Ala Leu Glu
 15 145 150 155 160

Thr Leu Lys Gln Ala Ala Lys Glu Ala Val Gln Glu Asp Gly Ala Glu
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20 Ser Ile Val Leu Gly Cys Ala Gly Met Val Gly Phe Ala Arg Gln Leu
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Asn Ser Tyr Gln Lys Pro Thr Glu Lys Gln Tyr Leu
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335

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40 cgg gat atc agc agt ggg gcg ggc cac gac tcg atg ttc atc gcc cag 1104
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 45 Val Thr Asp Val Gly Met Val Phe Val Pro Ser Arg Ala Gly Arg Ser
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10	Tyr	Thr	Pro	Glu	His	Ala	Ala	Ala	Arg	Glu	Thr	Leu	Ile	Ala	Ala	Met	35	40	45	
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	Ser	His	Phe	Asp	Ser	Val	Arg	Asn	Gly	Gly	Met	Phe	Asp	Gly	Thr	Ala	85	90	95	
20	Gly	Val	Val	Cys	Ala	Leu	Glu	Ala	Ala	Arg	Val	Met	Leu	Glu	Asn	Gly	100	105	110	
25	Tyr	Val	Asn	Arg	His	Pro	Phe	Glu	Phe	Ile	Ala	Ile	Val	Glu	Glu	Glu	115	120	125	
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40	His	Ile	Glu	Gln	Gly	Pro	Ile	Leu	Glu	Gln	Glu	Gln	Ile	Glu	Ile	Gly	195	200	205	
	Val	Val	Thr	Ser	Ile	Val	Gly	Val	Arg	Ala	Leu	Arg	Val	Ala	Val	Lys	210	215	220	
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	Ala	Leu	Val	Pro	Ala	Ala	Leu	Met	Val	Arg	Glu	Val	Asn	Arg	Phe	Val	245	250	255	
50	Asn	Glu	Ile	Ala	Asp	Gly	Thr	Val	Ala	Thr	Val	Gly	His	Leu	Thr	Val	260	265	270	
55	Ala	Pro	Gly	Gly	Gly	Asn	Gln	Val	Pro	Gly	Glu	Val	Asp	Phe	Thr	Leu	275	280	285	
	Asp	Leu	Arg	Ser	Pro	His	Glu	Glu	Ser	Leu	Arg	Val	Leu	Ile	Asp	Arg	290	295	300	

43

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5 Asp Val Asp Glu Phe Phe Asn Leu Ser Pro Val Gln Leu Ala Pro Thr
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55 tct gcg gcc gtg gga ggc atc acc acc atc atc gag atg ccg atc acc 288
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 Phe Pro Pro Thr Thr Thr Leu Asp Ala Phe Leu Glu Lys Lys Lys Gln

14

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5	gcg ggg cag cgg ttg aaa gtt gac ttc gcg ctc tat gga ggt gga gtg Ala Gly Gln Arg Leu Lys Val Asp Phe Ala Leu Tyr Gly Gly Gly Val 115 120 125			384
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25	ggg tca gtc atc gtg gtt cat gcc gag aat gaa acg atc att caa gcg Gly Ser Val Ile Val Val His Ala Glu Asn Glu Thr Ile Ile Gln Ala 180 185 190			576
30	ctc cag aag cag atc aag gcc gct ggc ggc aag gac atg gcc gcc tac Leu Gln Lys Gln Ile Lys Ala Ala Gly Gly Lys Asp Met Ala Ala Tyr 195 200 205			624
35	gag gca tcc caa cca gtt ttc cag gag aac gag gcc att cag cgt gcg Glu Ala Ser Gln Pro Val Phe Gln Glu Asn Glu Ala Ile Gln Arg Ala 210 215 220			672
40	ttg ctt ctg cag aaa gaa gcc ggc tgt cga ctg atc gtg ctt cac gtg Leu Leu Leu Gln Lys Glu Ala Gly Cys Arg Leu Ile Val Leu His Val 225 230 235 240			720
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65	gtc atc gac acc ctt gga tca gat cat ggc gga cat cct gtc gag gac Val Ile Asp Thr Leu Gly Ser Asp His Gly Gly His Pro Val Glu Asp 305 310 315 320			960
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Phe Pro Pro Thr Thr Thr Leu Asp Ala Phe Leu Glu Lys Lys Lys Gln
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16

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Gly Ser Val Ile Val Val His Ala Glu Asn Glu Thr Ile Ile Gln Ala
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15 Leu Gln Lys Gln Ile Lys Ala Ala Gly Gly Lys Asp Met Ala Ala Tyr
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20 Glu Ala Ser Gln Pro Val Phe Gln Glu Asn Glu Ala Ile Gln Arg Ala
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Leu Leu Leu Gln Lys Glu Ala Gly Cys Arg Leu Ile Val Leu His Val
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Gln Asp Val His Cys Glu Ser Gly Pro Gln Tyr Leu Asn Ile Thr Thr
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45 Gly Arg Leu Ser Leu Glu Arg Leu Val Glu Val Met Cys Glu Lys Pro
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50 Ala Lys Leu Phe Gly Ile Tyr Pro Gln Lys Gly Thr Leu Gln Val Gly
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Ser Asp Ala Asp Leu Leu Ile Leu Asp Leu Asp Ile Asp Thr Lys Val
385 390 395 400

55 Asp Ala Ser Gln Phe Arg Ser Leu His Lys Tyr Ser Pro Phe Asp Gly
405 410 415

Met Pr

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/55 C12N15/61 C12P13/00 C12P13/22 C12P41/00
C12N1/21 C12Q1/68

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N C12P C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

BIOSIS, MEDLINE, EPO-Internal, WPI Data, PAJ, STRAND, EMBL

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WATABE K ET AL: "IDENTIFICATION AND SEQUENCING OF A GENE ENCODING A HYDANTOIN RACEMASE FROM THE NATIVE PLASMID OF PSEUDOMONAS-SP STRAIN NS671" JOURNAL OF BACTERIOLOGY, vol. 174, no. 11, 1992, pages 3461-3466, XP000944037 ISSN: 0021-9193 page 3461, left-hand column, paragraph 2 -page 3463, right-hand column, paragraph 2; figures 3,4 page 3465, right-hand column page 3466, left-hand column, paragraphs 3-5	1-3,5,6, 11
Y	page 3465, right-hand column, paragraph 1 -page 3466, left-hand column, paragraph 1	1-3,5,6, 11,13
Y	page 3462, left-hand column, paragraph 5 -page 3462, right-hand column, paragraph 1	9
Y	page 3462 -/--	4

☒ Further documents are listed in the continuation of box C.

☐ Patent family members are listed in annex.

° Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- *&* document member of the same patent family

Date of the actual completion of the international search

11 December 2000

Date of mailing of the international search report

16.01.01

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+31-70) 340-3016

Authorized officer

Steffen, P

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>---</p> <p>GRIFANTINI RENATA ET AL: "Efficient conversion of 5-substituted hydantoins to D-alpha-amino acids using recombinant Escherichia coli strains." MICROBIOLOGY (READING), vol. 144, no. 4, April 1998 (1998-04), pages 947-954, XP002154848 ISSN: 1350-0872 page 949; figure 1 page 950, right-hand column, paragraph 2 -page 952, right-hand column, paragraph 1; table 4 page 953, left-hand column</p>	1-3,5,6, 11,13
X	<p>---</p> <p>SIEMANN MARTIN ET AL: "Characterization of serological properties of polyclonal antibodies produced against enzymes involved in the L-selective cleavage of hydantoin derivatives." BIOTECHNOLOGY LETTERS, vol. 15, no. 1, 1993, pages 1-6, XP000944249 ISSN: 0141-5492 page 1, paragraph 1 -page 2, paragraph 4 page 5; table 1</p>	14,16
Y	<p>---</p> <p>page 2; table 1</p>	4
X	<p>---</p> <p>WILMS BURKHARD ET AL: "Cloning, nucleotide sequence and expression of a new L-N-carbamoylase gene from Arthrobacter aurescens DSM 3747 in E. coli." JOURNAL OF BIOTECHNOLOGY, vol. 68, no. 2-3, 19 February 1999 (1999-02-19), pages 101-113, XP004164275 ISSN: 0168-1656 page 102, right-hand column, paragraph 4 -page 103, left-hand column, paragraph 1 page 109, left-hand column page 103 -page 104, left-hand column</p>	14,16
Y	<p>---</p>	9
X	<p>---</p> <p>BLATTNER FREDERICK R ET AL: "The complete genome sequence of Escherichia coli K-12." SCIENCE (WASHINGTON D C), vol. 277, no. 5331, 1997, pages 1453-1462, XP002069950 ISSN: 0036-8075 page 1454, right-hand column, paragraph 3</p> <p>---</p> <p>-/--</p>	15

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>DATABASE GENBANK 'Online! NCBI; Acc No: J01566, 8 February 1996 (1996-02-08) LEBOWITZ, J.: "Plasmid ColE1, complete genome" retrieved from NCBI, accession no. http://www.ncbi.nlm.nih.gov:80/ Database accession no. http://www.ncbi.nlm.nih.gov:80/Genbank/index.html XP002154850 the whole document</p>	15
P,X	<p>MAY OLIVER ET AL: "Inverting enantioselectivity by directed evolution of hydantoinase for improved production of L-methionine." NATURE BIOTECHNOLOGY, vol. 18, no. 3, March 2000 (2000-03), pages 317-320, XP002154849 ISSN: 1087-0156 page 318, right-hand column, paragraph 2 -page 319, left-hand column, paragraph 1 page 319, right-hand column, paragraph 3 -page 320, left-hand column, paragraph 1</p>	1-16
P,X	<p>WIESE ANJA ET AL: "Hydantoin racemase from <i>Arthrobacter aurescens</i> DSM 3747: Heterologous expression, purification and characterization." JOURNAL OF BIOTECHNOLOGY, vol. 80, no. 3, 2000, pages 217-230, XP000943983 ISSN: 0168-1656 page 220, right-hand column, paragraph 2 -page 222, left-hand column, paragraph 1; figures 2,4</p>	1-16
A	<p>WATABE K ET AL: "CLONING AND SEQUENCE OF THE GENES INVOLVED IN THE CONVERSION OF 5-SUBSTITUTED HYDANTOINS TO THE CORRESPONDING L AMINO ACIDS FROM THE NATIVE PLASMID OF PSEUDOMONAS-SP STRAIN NS671" JOURNAL OF BACTERIOLOGY, vol. 174, no. 3, 1992, pages 962-969, XP000944036 ISSN: 0021-9193 the whole document</p>	

INTERNATIONAL SEARCH REPORT

International application No.
PCT/EP 00/08473

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☒ Claims Nos.: 6,8,9,13 (partially)
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 6,8,9,13 (partially)

Present claims 6, 8, 9 and 13 relate to an extremely large number of possible methods. Support within the meaning of Article 6 PCT and/or disclosure within the meaning of Article 5 PCT is to be found, however, for only a very small proportion of the methods claimed. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Consequently, the search has been carried out for those parts of the claims which appear to be supported and disclosed, namely when the whole cell catalyst as referred to in claims 6, 8, 9 and 13 is restricted to a whole cell catalyst according to claim 1.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.